

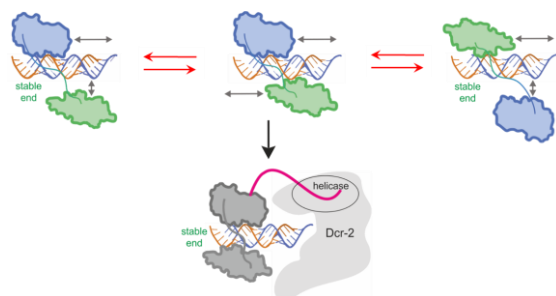
# Conformational Dynamics in Biomolecular Recognition & Optimization of NMR Experiments for Ultrahigh Magnetic Fields

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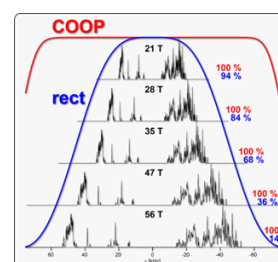
Protein-RNA interactions play essential roles in virtually all aspects of gene regulation, where single- or doublestranded RNAs are recognized by RNA binding proteins (RBPs). Most eukaryotic RBPs are multi-domain proteins that comprise multiple structural domains to mediate protein-RNA or protein-protein recognition often employing avidity, cooperativity and dynamic interactions [1-5].



We employ integrative structural biology to unravel the molecular mechanisms involving these regulatory RNP (ribonucleoprotein) complexes. For these studies, solution NMR-spectroscopy and SAXS/SANS provide unique information on functionally important dynamics and are combined with X-ray crystallography and electron microscopy to elucidate the structural mechanisms and dynamics of regulatory RNPs.

Recent progress will be presented that highlight the role of conformational dynamics and population shifts in molecular recognition of single- and double-stranded RNA by RBPs involved in splicing regulation and microRNA biogenesis.

Current progress will also be presented on the development of radiofrequency pulses and pulse elements using optimal control methods to enable NMR spectroscopy at ultrahigh magnetic fields (>1 GHz) which we pursue in collaboration with the Glaser lab at TUM. We have previously developed an ultrabroadband decoupling scheme with minimal decoupling artifacts at modest *rf* power [6]. We are now implementing cooperative pulses to achieve broadband excitation and chemical shift evolution in common biomolecular NMR pulse sequences with improved performance at current and future available magnetic fields.



## References

1. Supekar S, Papageorgiou AC, Gemmecker G, Peltzer R, Johansson MP, Tripsianes K, Sattler M\*, Kaila VRI\* Conformational Selection of Dimethylarginine Recognition by the Survival Motor Neuron Tudor Domain. (2018) **Angew Chem Int Ed Engl** doi: 10.1002/anie.201708233
2. Sonntag M, Jagtap PKA, Simon B, Appavou MS, Geerlof A, Stehle R, Gabel F, Hennig J, and Sattler M Segmental, Domain-Selective Perdeuteration and Small-Angle Neutron Scattering for Structural Analysis of Multi-Domain Proteins. (2017) **Angew Chem Int Ed Engl**. doi: 10.1002/anie.201702904
3. Tants JN, Fesser S, Kern T, Stehle R, Geerlof A, Wunderlich C, Juen M, Hartlmüller C, Bottcher R, Kunzelmann S, Lange O, Kreutz C, Forstemann K\*, and Sattler M\* *Molecular basis for asymmetry sensing of siRNAs by the Drosophila Loqs-PD/Dcr-2 complex in RNA interference.* (2017) **Nucleic Acids Res.** doi: 10.1093/nar/gkx886
4. Mourao A, Bonnal S, Soni K, Warner L, Bordonne R, Valcarcel J, and Sattler M *Structural basis for the recognition of spliceosomal SmN/B/B' proteins by the RBM5 OCRE domain in splicing regulation.* (2016) **Elife** 5, e14707. doi: 10.7554/eLife.14707
5. Voith von Voithenberg L, Sanchez-Rico C, Kang HS, Madl T, Zanier K, Barth A, Warner LR, Sattler M, and Lamb DC *Recognition of the 3' splice site RNA by the U2AF heterodimer involves a dynamic population shift.* (2016) **Proc Natl Acad Sci U S A** 113, E7169-E7175. doi: 10.1073/pnas.1605873113
6. Schilling F, Warner LR, Gershenson NI, Skinner TE, Sattler M, Glaser SJ. Next-Generation Heteronuclear Decoupling for High-Field Biomolecular NMR Spectroscopy. (2014) **Angew Chem Int Ed Engl**. 53, 4475–4479. doi: 10.1002/anie.201400178